

# The effects of UV radiation on anodic wave of human serum

Gülten Atun<sup>a,\*</sup>, Hilal Nesibe Haznedaroğlu<sup>b</sup>

<sup>a</sup> *Istanbul University, Department of Chemistry, Avcılar, 34320, Turkey*

<sup>b</sup> *Çekmece Nuclear Research and Training Center P. Box 1 Airport 34 481 Istanbul, Turkey*

Received 28 February 2007; received in revised form 8 November 2007; accepted 18 November 2007

Available online 23 November 2007

## Abstract

Polarographic anodic oxidation wave that can be correlated to total antioxidant capacity (TAC) in Human serum corresponds to  $2\text{H}^+$  and  $2e$  process. The limiting current of the wave, which is proportional to TAC, is strongly influenced by UV light intensity and irradiation time. A mechanism has been proposed to explain the effects of UV on anodic wave related to antioxidant behavior. Results show that decomposition processes follow to the excitation a transitory product at three different light intensities at 340 nm. The number of decomposition products increase with increasing light intensity. The rate of oxygen radical capture ability of the serum antioxidants might be estimated by comparing the changes of anodic wave in the absence and presence of naturally dissolved oxygen.

© 2007 Elsevier B.V. All rights reserved.

**Keywords:** Human serum; Anodic wave; UV radiation; Antioxidant; Reaction rate

## 1. Introduction

Antioxidants protect cells against the damaging effects of reactive oxygen species consisting of free radicals such as  $\text{O}^{\cdot-}$ ,  $\text{OH}^{\cdot}$ , and other non-radical oxygen derivatives such as hydrogen peroxide ( $\text{H}_2\text{O}_2$ ), hypochlorous acid ( $\text{HOCl}$ ) and singlet Oxygen ( $^1\text{O}_2$ ). An imbalance between antioxidants and reactive oxygen species results in oxidative stress, which has been linked to cancer, aging, atherosclerosis, ischemic injury, inflammation and neurodegenerative diseases [1]. Biological fluids contain numerous compounds having antioxidant activity, including urate, ascorbate, bilirubin, and thiols in the aqueous phase and  $\alpha$ -tocopherol, carotenoids, and flavonoids in the lipid phase [2]. Tryptophan, tyrosine, cysteine and homocysteine also show antioxidant ability [3]. Cooperation of all the different antioxidants provides greater protection against attack by reactive oxygen radicals, than any single compound alone. Thus, the TAC may give more relevant biological information compared to that obtained by the measurement of individual components, as it considers the cumulative effect of all antioxidants

present in plasma and body fluids. Because of difficulty in measuring each antioxidant component separately and interactions among antioxidants, methods have been developed to assess the TAC of serum or plasma such as Trolox-equivalent antioxidant capacity (TEAC), the oxygen radical absorbance capacity (ORAC), and the ferric reducing ability of plasma (FRAP), the crocin bleaching and the CUPRAC assays as well as high-throughput fluorescence screening, enhanced chemiluminescence and cyclic voltammetry (CV) techniques are commonly used and have been extensively evaluated [1–13].

By using the CV technique, the reducing power of antioxidants is characterized from the peak potential ( $E_p$ ) or inflection point of current wave (half wave potential,  $E_{1/2}$ ) of the anodic peak current ( $I_p$ ), which provides information on the ability of antioxidants to donate electrons. The TAC in plasma and other biological fluids has been determined from the anodic peak current using cyclic voltammetry (CV) [10–13]. The anodic peak having the  $E_p$  of  $\sim 0.45$  V and  $E_{1/2}$  of  $\sim 0.3$  V in human plasma has been attributed to TAC of the plasma [11].

Some compounds such as vitamin C, uric acid and bioflavonoids, which contribute largely to TAC level also give a polarographic wave around similar potentials [14–19]. These biomolecules can absorb UV-A (320–400 nm) and UV-B (280–

\* Corresponding author. Tel.: +90 212 4737031; fax: +90 212 4737180.

E-mail address: [gultena@istanbul.edu.tr](mailto:gultena@istanbul.edu.tr) (G. Atun).

320 nm) radiation leading to intermediates that can remove photodynamically generated reactive oxygen [17–20]. The electrochemical oxidation of uric acid has been studied in NaCl and phosphate buffers [20]. The kinetic investigations show that a UV-absorbing intermediate is produced during electrooxidation of uric acid (7,9-dihydro-1*H*-purine-2,6,8-(3*H*)-trione). On the basis of pseudo first order rate constants observed in different media, it has been concluded that the diimine species formed from the  $2e, 2H^+$  oxidation of uric acid is attacked by water to give diol, which then decomposes in a series of reactions to give allantoin as the major product. Several investigators have also studied action of UV radiation on human blood serum [21–24].

In this study, a mechanism has been proposed for antioxidant activity in human serum by following the changes of polarographic anodic wave during UV irradiation.

## 2. Materials and methods

The freshly taken blood samples from healthy volunteers were centrifuged at 3000 rpm, by using Hettich centrifuge and the serum was separated as supernatant liquid. A  $0.075\text{ cm}^3$  of the serum was added into  $25\text{ cm}^3$  of  $0.1\text{ M KNO}_3$  solution served as a supporting electrolyte. The solution pH was 8.08. This aliquot was placed in a Pyrex polarographic cell with a thermostatic jacket covered with carbon black having a small circular area of  $1.0\text{ cm}^2$  to irradiate the reaction mixture. A two-electrode system was used for obtaining a polarographic wave. Double distilled mercury dropping electrode (DME) was used as a working electrode and a saturated calomel electrode (SCE) served as the reference. The SCE was separated from the cell by a glass frit. The dropping time of DME was adjusted as 2.7 s. per drop. The solution was deaerated by bubbling nitrogen gas for 15 min. Polarograms were recorded using a Radiometer Copenhagen PO-4 DC Polarograph. Thus, a reproducible well-defined anodic oxidation wave was obtained at a scan rate of  $13.3\text{ mV s}^{-1}$ .

The solutions were irradiated using a high-pressure mercury vapor lamp (Edison 400 HQL) as UV source. Monochromatic UV light was obtained using a Kodak glass filter of 340 nm. Absorbing light intensities ( $I_a$ ) were adjusted by changing the distance of the UV source to the system. The values of  $I_a$  were determined as  $0.66 \times 10^{15}$ ,  $1.50 \times 10^{15}$  and  $3.70 \times 10^{15}\text{ quanta s}^{-1}\text{ cm}^{-2}$  using ferrioxalate actinometer. The anodic waves were recorded in regular time intervals during irradiation at varying light intensities. During irradiation, the temperature of the solution was kept constant at 298 K.

Representative polarograms recorded at various irradiation time intervals at  $0.66 \times 10^{15}\text{ quanta s}^{-1}\text{ cm}^{-2}$  light intensity are shown in Fig. 1.

The experiment was also conducted in the presence of naturally dissolved oxygen at medium light intensity of  $1.50 \times 10^{15}\text{ quanta s}^{-1}\text{ cm}^{-2}$ .

## 3. Results and discussion

Serum solution yields a reproducible well-developed anodic wave at the half wave potential of 0.28 V. The limiting current

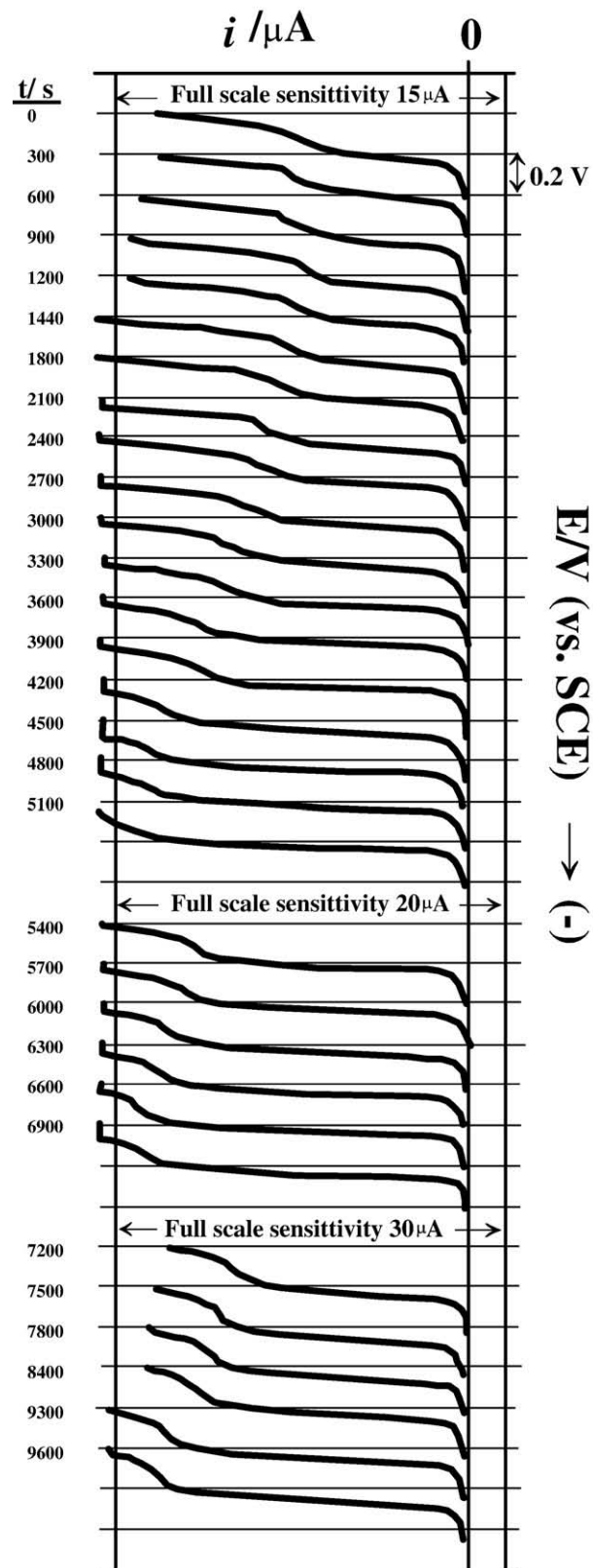


Fig. 1. Representative polarograms for irradiation with UV light of low intensity at 340 nm wavelengths depending on time (Starting potential is 0.5 V and full scale sensitivity of diffusion currents is shown at the beginning of each series).

of the wave is  $\sim 5 \mu\text{A}$  in deaerated solution. This value was measured for an anodic wave at  $t=0$  when there is no irradiation. Since the residual current increases with the applied e.m.f the initial point of the diffusion current is determined from the intersection point of extrapolated lines of residual- and diffusion currents. The true limiting current is determined by measuring the distance between the total diffusion current and its initial point. The estimated uncertainty of the measured limiting currents is  $\pm 3\%$ .

An anodic wave corresponding to a reversible electrode process can be analyzed according to following Heyrovsky-Ilkovich equation [25,26]:

$$E_{\text{d.e.}} = E_{1/2} + \frac{0.059}{n} \log \frac{(i_d - i)}{i} \quad (1)$$

where  $E_{\text{d.e.}}$  and  $i$  are corresponding voltage and current values at any point on the wave,  $E_{1/2}$  is half wave potential,  $i_d$  is limiting current and  $n$  the number of electrons transferred in the electrode process. It is evident from the Eq. (1) a plot of  $E_{\text{d.e.}}$  vs.  $\log (i_d - i)/i$  plot should produce a straight line with a slope  $0.059/n$  at 298 K, and potential where the log term becomes zero should be the half wave potential. If the electrode reaction is reversible but the reaction product formed by electrolysis is unstable and is irreversibly transformed another compound, the net reaction becomes irreversible. In such a case, it has been theoretically proved that the shape of the anodic wave corresponding to  $2e, 2H^+$  process at a given pH is identical with that of a reversible reaction, which involves the transfer of two electrons [25,26]. Similar mechanism has been proposed for the anodic waves of some antioxidants such as ascorbic acid [14], hesperidin [17], rutin [18], morin [19] and uric acid [20].

By plotting the values of  $E_{\text{d.e.}}$  vs.  $\log (i_d - i)/i$  a straight line was obtained with a slope of 0.031 (not shown here,  $r^2=0.9998$ ), in good agreement with theoretical slope of 0.0296 corresponding to  $2e, 2H^+$  process. The value of  $E_{1/2}$  of 0.28 V is not oxidation potential of the reversible electrode reaction it also involves the rate constant of the irreversible reaction.

Consequently, the anodic wave of serum can be attributed to many species present in human serum that are simultaneously oxidized with the same mechanism and the electrode reactions of these compounds may be generalized in the following form [14,17–20,25,26]:



Here, AO denotes antioxidant and DA is its dehydro-form, and  $k$  is the rate constant.

Limiting current of the anodic wave ( $i_d$ ) is proportional to  $[DA]$  concentration according to Ilkovic equation [25]. The changes in limiting diffusion currents depending on irradiation time at varying light intensities were depicted in Fig. 2. The values of  $i_d$  show an increase at earlier irradiation time at the light intensity of  $0.66 \times 10^{15} \text{ quanta s}^{-1} \text{ cm}^{-2}$  but they slow down as the time increases (triangles). The increase in  $i_d$  values for medium light intensity is higher than at the lowest intensity

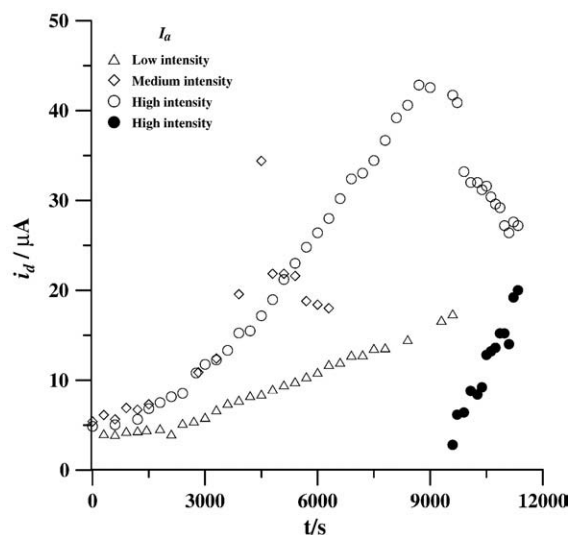


Fig. 2. The influences of UV radiation with different intensities on limiting current of anodic wave of Human serum.

initially whereas a gradual decrease is observed following a maximum value (diamonds).

A slower increase is observed in the  $i_d$  values at the highest absorbing intensity of  $3.70 \times 10^{15} \text{ quanta s}^{-1} \text{ cm}^{-2}$  and the maximum value of  $i_d$  is higher than that of medium light intensity (open circles). When  $i_d$  values start to decrease another polarographic wave is observed at the half wave potential of 0.38 V. This wave also increases by irradiation (solid circles).

In order to explain the effects of UV light on anodic wave, which can be correlated to antioxidant behavior, following mechanism has been proposed to account for the observed results based on electrode process [14,17–20,25,26]. Each step can play a role in the overall mechanism, depending on light intensity and irradiation time.



Here, the Eqs. (4)–(6) are elementary steps of electrode process while the Eqs. (7)–(11) are decomposition steps of the

dehydro-form of antioxidants which can give B, C, D, E, F, G, H, I, J and K decomposition products. The rate constant for a respective rate process is shown on the arrow.

When serum is irradiated by UV light, the changes in the DA concentration with irradiation time may be written as follows:

$$\frac{d[DA]}{dt} = k_3[DA^*] - k_4[DA] - k_5[DA] - k_6[DA] - k_7[DA] - k_8[DA] \quad (12)$$

where, the brackets indicate the concentrations of the various species. The concentration of the transitory product  $[DA^*]$  in Eq. (12) can be determined from the stationary or steady state principle [27]:

$$k_1 I_a = k_2[DA^*] + k_3[DA^*] \quad (13)$$

Consequently,

$$[DA^*] = \frac{k_1 I_a}{k_2 + k_3} \quad (14)$$

The rate equation can be obtained in terms of the measurable  $[DA]$  concentration by substitution of  $[DA^*]$  in Eq. (12):

$$\frac{d[DA]}{dt} = \frac{k_1 k_3 I_a}{k_2 + k_3} - k_4[DA] - k_5[DA] - k_6[DA] - k_7[DA] - k_8[DA] \quad (15)$$

This equation is rearranged and integrated:

$$\int \frac{d[DA]}{[DA]} = \int \left( \frac{k_1 k_3 I_a}{k_2 + k_3} \times \frac{1}{[DA]} - k_4 - k_5 - k_6 - k_7 - k_8 \right) dt \quad (16)$$

$$\ln[DA] = \left( \frac{k_1 k_3 I_a}{k_2 + k_3} \times \frac{1}{[DA]} - k_4 - k_5 - k_6 - k_7 - k_8 \right) t + I \quad (17)$$

The value of integral constant,  $I$ , is calculated as:

$$I = \ln[DA]_0 \quad (18)$$

for initial irradiation time ( $t=0$ ).

$$\ln \frac{[DA]}{[DA]_0} = \left( \frac{k_1 k_3 I_a}{k_2 + k_3} \times \frac{1}{[DA]} - k_4 - k_5 - k_6 - k_7 - k_8 \right) t \quad (19)$$

$$k' = \frac{k_3}{k_2 + k_3} \quad (20)$$

$$\ln \frac{[DA]}{[DA]_0} = \left( \frac{k_1 k' I_a}{[DA]} - k_4 - k_5 - k_6 - k_7 - k_8 \right) t \quad (21)$$

Here,  $[DA]_0$  and  $[DA]$  are concentrations of dehydro product at initial and irradiation time  $t$ , respectively. The ratio of  $[DA]/[DA]_0$  corresponds to the ratio of limiting current at irradiation time  $t$  to that of its initial value. The values of  $k_1$ ,  $k_4$ ,  $k_5$ ,  $k_6$ ,  $k_7$  and  $k_8$  could be determined from the slopes of  $\ln[DA]/[DA]_0$  vs.

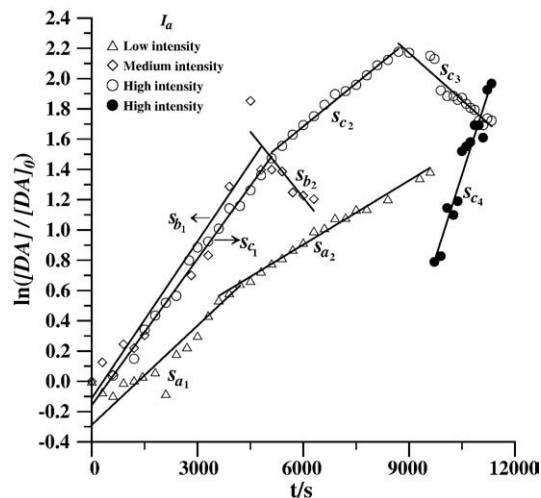


Fig. 3. The plot for determination of rate constants of photochemical reactions depending on irradiation time and light intensity ( $s$  denotes the slope of the straight line, subscripts a, b and c represent low, medium and high light intensity, respectively.).

$t$  plot in Fig. 3 for a known  $I_a$  and the standard concentration of  $[DA]=1 \text{ mol dm}^{-3}$ .

### 3.1. Determination of the rate constants

The rate constants can be determined by analyzing of  $\ln [DA]/[DA]_0$  vs.  $t$  curves at different light intensity and irradiation times as follows:

#### 3.1.1. Low light intensity

When serum solution is irradiated by UV light with low intensity  $\ln[DA]/[DA]_0$  vs.  $t$  plot gives two straight lines with decreasing slopes of  $s_{a1}$  and  $s_{a2}$ , respectively. By assuming the slope of the second straight line corresponds to rate constant ( $k_4$ ) of decomposition reaction in Eq. (7) slope of the first straight line can be derived from Eqs. (4)–(7) as follows:

$$s_a = \frac{k_1 k_3 I_a}{k_2 + k_3} - k_4 \quad (22)$$

Since electrode reaction becomes irreversible when irradiated by UV light  $k_2$  can be neglected. Thus Eq. (22) can be rearranged as follows:

$$s_{a1} = \frac{k_1 k_3 I_a}{k_3} - k_4 \quad (23)$$

$$s_{a1} = k_1 I_a - k_4 \quad (24)$$

The value of  $k_4$  is calculated as  $1.40 \times 10^{-4} \text{ s}^{-1}$  from the slope of the second straight line ( $r=0.99$ ). In the kinetic calculations the unit of  $I_a$  is converted from quanta  $\text{s}^{-1} \text{ cm}^{-2}$  into quanta  $\text{mol dm}^{-3} \text{ s}^{-1}$  by dividing it by Avogadro's number and the volume of the solution. Therefore, the value of  $k_1$  is calculated as  $7.69 \times 10^3 \text{ s}^{-1}$  by using the  $I_a$  value of  $4.38 \times 10^{-8} \text{ mol quanta dm}^{-3} \text{ s}^{-1}$  and the  $s_{a1}$  value of  $1.97 \times 10^{-4} \text{ s}^{-1}$  ( $r=0.96$ ).



### 3.1.2. Medium light intensity

As shown in Fig. 3, two straight lines are observed the first having a positive slope ( $s_{b_1}$ ) and the latter negative ( $s_{b_2}$ ) depending on increasing irradiation time at medium light intensity. These results suggest that reaction mechanism follows the order of Eqs. (4)–(8) and the slopes of the straight lines correspond to:

$$s_{b_1} = k_1 I_a - k_4 - k_5 \quad (25)$$

$$s_{b_2} = -k_5 \quad (26)$$

The value of  $k_5$  is calculated from the observed slope of the second straight-line as  $2.90 \times 10^{-4} \text{ s}^{-1}$  ( $r=0.94$ ). The  $s_{b_1}$  value is computed as  $3.37 \times 10^{-4} \text{ s}^{-1}$  by using the known value of  $I_a$  of  $9.97 \times 10^{-8} \text{ mol quanta dm}^{-3} \text{ s}^{-1}$  and  $k_1$ ,  $k_4$  and  $k_5$  values. This value is in agreement with the observed slope of the first straight line of  $3.45 \times 10^{-4} \text{ s}^{-1}$  ( $r=0.97$ ).

### 3.1.3. High light intensity

The  $\ln[\text{DA}]/[\text{DA}]_0$  vs.  $t$  plot derived from the main polarographic wave at  $E_{1/2}$  of 0.28 V gives three straight lines with two positive ( $s_{c_1}$  and  $s_{c_2}$ ) and one negative ( $s_{c_3}$ ) slopes. At longer irradiations, another straight line having a positive slope ( $s_{c_4}$ ) corresponding to  $E_{1/2}$  of 0.38 V is obtained when the main wave starts to decrease. The observed slope of the first straight line is well consistent with the calculated slope by assuming following equations:

$$s_{c_1} = k_1 I_a - k_4 - k_5 - k_6 - k_7 - k_8 \quad (27)$$

$$s_{c_2} = k_6 \quad (28)$$

$$s_{c_3} = -k_7 \quad (29)$$

$$s_{c_4} = k_8 \quad (30)$$

The  $s_{c_1} < s_{b_1}$  values suggest that additional decomposition reactions occur at the highest light intensity of  $2.46 \times 10^{-7} \text{ mol}$

$\text{quanta dm}^{-3} \text{ s}^{-1}$ . The rate constants correspond to additional decomposition reactions in Eqs. (9)–(11) could be calculated from the slopes of the straight lines according to Eqs. (28)–(30). The value of the rate constant  $k_6$  is  $1.90 \times 10^{-4} \text{ s}^{-1}$  ( $r=1.00$ ). The reaction products generated according to Eq. (10) are not polarographically active and the rate constant  $k_7$  can be calculated from the negative slope of the third straight line as  $2.10 \times 10^{-4} \text{ s}^{-1}$  ( $r=0.95$ ). On the other hand, a polarographically active product can be followed only after the main wave starts to decrease. The rate constant  $k_8$ , which corresponds to Eq. (11), can be determined from the positive slope of the fourth straight line as  $7.30 \times 10^{-4} \text{ s}^{-1}$  ( $r=0.98$ ). Using the known values of the constants involved in Eq. (27), the slope of the first straight line is calculated as  $3.31 \times 10^{-4} \text{ s}^{-1}$ . This value is in accordance with its observed value of  $3.22 \times 10^{-4} \text{ s}^{-1}$  ( $r=1.00$ ).

### 3.2. Effect of UV irradiation on anodic wave in the presence of oxygen

The attack of antioxidants on oxygen radical species may be better understood by comparing the behavior of anodic wave of serum in the presence of naturally dissolved oxygen to in its absence. A comparison of the limiting currents for irradiations with medium UV light intensity is presented in Fig. 4. As it is seen from Fig. 4, the height of anodic wave in the presence of oxygen is lower than that of its absence under dark conditions (i.e., irradiation time  $t=0$ ). This suggests that antioxidants present in Human serum are loosely bounded with molecular oxygen, which can be interrupted during nitrogen bubbling under pressure. After removal of oxygen the wave height increases by about two folds.

When serum irradiated under oxic conditions the increase of the wave height is very slow suggesting that light-induced antioxidant species capture the reactive oxygen species, which are produced upon UV radiation. The rate of oxygen radical capture of the serum antioxidants can be estimated as  $2.96 \times 10^{-4} \text{ s}^{-1}$  from the difference of the slopes of  $\ln[\text{DA}]/[\text{DA}]_0$  vs.  $t$  plots obtained in the absence and in the presence of oxygen.

## 4. Conclusions

Polarographic technique has provided valuable information about electron transfer mechanism and decomposition reactions of serum antioxidants under UV exposure without any separation procedure.

The polarographic anodic wave of human serum related to TAC has been interpreted on the basis of a mechanism involving a reversible electrode reaction followed by the irreversible conversion of an unstable intermediate to its stable form. When serum is irradiated by UV light reversible electrode process becomes irreversible and decomposition products are generated depending on light intensity and irradiation time.

The reactions initiated with different light intensities are completed at different irradiation time which are reflected by the straight lines having different slopes on  $\ln[\text{DA}]/[\text{DA}]_0$  vs.  $t$  plot. Results show that one, two and five decomposition reactions appear by irradiation at low, medium and high intensities,

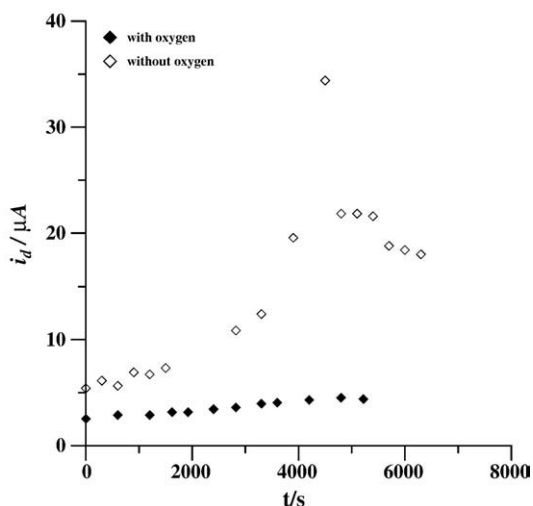


Fig. 4. A comparison of the limiting currents of anodic waves irradiated at the medium light intensity in the presence and in the absence of oxygen.

respectively. The rate constant  $k_1$  of the irreversible electrode process and  $k_4$  corresponding to the first decomposition reaction can be obtained by irradiations at low intensity. At medium intensity, the first decomposition is completed at earlier time of irradiations and rate constant of the second decomposition ( $k_5$ ) can be calculated from the slope of the second straight line. Similarly, the first and the second decomposition processes are completed at initial irradiations with high intensity UV light and the corresponding rate constants of three decompositions ( $k_6$ – $k_8$ ) are calculated from the slopes of the three straight lines, which appear depending on irradiation time.

A comparison of the anodic waves irradiated under oxic and anoxic conditions confirmed that light-induced antioxidant species capture light-induced oxygen species, which are highly reactive and damaging to humans at the molecular level.

## References

- [1] B. Mayer, M. Schumacher, H. Brandstatter, F.S. Wagner, A. Hermetter, High-throughput fluorescence screening of antioxidative capacity in human serum, *Anal. Biochem.* 297 (2001) 144–153.
- [2] I.S. Young, Measurement of total antioxidant capacity, *J. Clin. Pathol.* 54 (2001) 339, doi:10.1136/jcp.54.5.339 [jcp.bmjournals.com](http://jcp.bmjournals.com).
- [3] E. Meucci, M.C. Mele, Amino acids and plasma antioxidant capacity, *Amino Acids* 12 (1997) 373–377.
- [4] G. Cao, R.L. Prior, Comparison of different analytical methods for assessing total antioxidant capacity of human serum, *Clin. Chem.* 44 (1998) 1309–1315.
- [5] C.C. Wang, C.Y. Chu, K.O. Chu, K.W. Choy, K.S. Khaw, M.S. Rogers, C.P. Pang, Trolox-equivalent antioxidant capacity assay versus oxygen radical absorbance capacity assay in plasma, *Clin. Chem.* 50 (2004) 952–954.
- [6] I.F.F. Benzie, J.J. Strain, The ferric reducing ability of plasma (FRAP) as a measure of 'antioxidant power': The FRAP assay, *Anal. Biochem.* 239 (1996) 70–76.
- [7] K. Marilena, A. Nistikaki, V. Tsaousis, N. Maliaraki, G. Notas, E. Castanas, A new automated method for the determination of the Total Antioxidant Capacity (TAC) of human plasma, based on the crocin bleaching assay, *BMC Clin. Pharmacol.* 2 (2002), doi:10.1186/1472-6890-2-3 <http://www.biomedcentral.com>.
- [8] R. Apak, K. Güçlü, M. Özyürek, S.E. Karademir, M. Altun, Total antioxidant capacity assay of human serum using copper (II)-neocuproine as chromogenic oxidant: the CUPRAC method, *Free Radic. Res.* 39 (2005) 949–961.
- [9] M. Ryan, L. Grayson, D.J. Clarke, The total antioxidant capacity of human serum measured using enhanced chemiluminescence is almost completely accounted for by urate, *Ann. Clin. Biochem.* 34 (1997) 688–689.
- [10] C. Rice-Evans, N.J. Miller, Total antioxidant status in plasma and body fluids, *Methods Enzymol.* 234 (1994) 279–293.
- [11] J. Psotová, J. Zahálková, J. Hrbáč, V. Šimánek, J. Bartek, Determination of total antioxidant capacity in plasma by cyclic voltammetry. Two case reports, *Biomed. Papers* 145 (2001) 81–83.
- [12] R. Kohen, E. Vellaichamy, J. Hrbac, I. Gati, O. Tirosh, Quantification of the overall reactive oxygen species scavenging capacity of biological fluids and tissues, *Free Radic. Biol. Med.* 28 (2000) 871–879.
- [13] R. Kohen, E. Beit-Yannai, E.M. Berry, O. Tirosh, Overall low molecular weight antioxidant activity of biological fluids and tissues by cyclic voltammetry, *Methods Enzymol.* 300 (1999) 285–296.
- [14] D.M.H. Kern, The polarographic oxidation potential of ascorbic acid, *J. Am. Chem. Soc.* 76 (1954) 1011–1015.
- [15] J.J. Ruiz, A. Aldaz, M. Domínguez, Mechanism of L-ascorbic acid oxidation and dehydro-L-ascorbic acid reduction on a mercury electrode. I. Acid medium, *Can. J. Chem./Rev. Can. Chim.* 55 (1977) 2799–2806.
- [16] A. Yu, H. Zhang, H. Chen, Catalytic oxidation of uric acid at the polyglycine chemically modified electrode and its trace determination, *Analyst* 122 (1997) 839–841.
- [17] S. Baykut, G. Atun, Investigation of rutins photooxidation by polarographic method, *Chim. Acta Turc.* 17 (1990) 339–353.
- [18] S. Baykut, G. Atun, Investigation of photochemical oxidation of hesperidin and determination of the number of lost electron in the reaction by polarographic method, *Chim. Acta Turc.* 18 (1989) 365–374.
- [19] T. Şişmanoğlu, G. Atun, The effect of UV radiation on anodic wave of morin, *Chim. Acta Turc.* 28 (2000) 11–14.
- [20] R.N. Goyal, A. Mittal, D. Agarwal, Electrochemical oxidation and kinetics of decay of UV-absorbing intermediates uric acid oxidation at pyrolytic graphite electrodes, *Can. J. Chem.* 72 (1994) 1668–1675.
- [21] V.Yu. Kulikov, V.P. Kaznatcheev, L.I. Kolesnikova, Action of UV radiation on human blood serum under normal and pathological conditions, *Bull. Exp. Biol. Med.* 83 (1977) 497–499.
- [22] N. Kollias, I. Baqer, Sadiq, R.M. Sayre, In vitro and in vivo ultraviolet induced alterations of oxy- and deoxyhemoglobin, *Photochem. Photobiol.* 56 (1992) 223–227.
- [23] L.S. Demma, J.M. Salhanry, Subunit inequivalence in superoxide anion formation during photo-oxidation of human oxyhemoglobin, *J. Biol. Chem.* 254 (1979) 4532–4535.
- [24] U. Zelck, U. Karnstedt, F.W. Gansicke, S. Wiesner, P. Lange, Mechanism of action of ultraviolet irradiation of blood: on the possible role of the anti-oxidative enzyme, VASA, *Suppl.* 32 (1991) 365–369.
- [25] I.M. Kolthoff, J.J. Lingane, *Polarography*, Intersci. Publ., New York, 1952.
- [26] L.I. Smith, I.M. Kolthoff, S. Wawzonek, P.M. Ruoff, *The Chemistry of Vitamin E. XXIX. Studies of the Behavior of Compounds Related to Vitamin E at the Dropping Mercury Electrode*, 1941, pp. 1018–1024.
- [27] S.H. Maron, J.B. Lando, *Fundamentals of Physical Chemistry*, Macmillan Publ. Co., Inc., New York, 1974.